

# Role of Bcl-2, p53, and Ki-67 expression in basal cell carcinoma and their association with aggressive and non-aggressive histological phenotypes

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## Abstract

**Introduction:** There is increasing evidence that immunohistochemical expression of p53, Ki-67, and Bcl-2 is associated with aggressive (aBCC) and less aggressive (nBCC) histological subtypes and may have a prognostic role.

**Aim:** To investigate the clinicopathological features and immunohistochemical expressions of p53, Ki-67, and Bcl-2 in cutaneous basal cell carcinoma focusing on histological subtypes. Their roles and possible interactions in the development and progression of BCC are discussed.

**Material and methods:** A total of 50 BCC samples from 50 patients from Western Mexico between June 2018 and June 2019 were included. Paraffin-embedded samples were immunostained with p53, Ki-67, and Bcl-2 antibodies. Semi-quantitative analysis was performed to determine the intensity and positivity of immunostained cells. Parametrical and non-parametrical tests were performed according to the sample's distribution.

**Results:** Samples included 21 nBCC and 29 aBCC. The statistical analysis showed statistical association when grouped as non-aggressive and aggressive subtypes for p53 ( $p = 0.04$ ) and Bcl-2 ( $p < 0.01$ ). An inverse negative correlation was found between age and Bcl-2 expression. No statistical association was found between Ki-67 immunoreactivity and any of the other variables.

**Conclusions:** We found that a high expression of Bcl-2 and a low expression of p53 was associated with more indolent histopathological features of BCC and therefore better outcomes. These findings suggest that examination of p53 and Bcl-2 expression in BCC patients may provide valuable prognostic information. These biomarkers may play a role in the development and progression of some cases of BCC.

**Key words:** Ki-67 antigen, tumour suppressor protein p53, prognosis, BCL2 protein, human, carcinoma, basal cell.

## Introduction

Basal cell carcinoma (BCC) has a slow growth rate, minimal soft tissue invasiveness, and a high cure rate. Occasionally, BCC can behave aggressively, invading deep tissues, and potentially having metastatic behaviour. Nodular and superficial BCC subtypes are classically acknowledged as non-aggressive or less aggressive BCCs (nBCC), while more aggressive BCCs (aBCC) include the

following patterns: metatypical, micronodular and infiltrative or morpheaform [1].

The association between the prognosis of solid tumours and the p53, Ki-67, and Bcl-2 biomarker expression has not been fully elucidated. The tumour suppressor p53 is the most frequently mutated gene in human cancers, and its inactivation is the second most frequent event following upregulation of the Hedgehog signalling pathway

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in BCC [2]. Several studies on p53 expression in BCCs reported a significantly greater expression of p53 in the aggressive groups, stating that p53 immunopositivity is an important prognostic factor for these tumours [3].

Expression of the nuclear protein Ki-67 is associated with cell proliferation. It has been used as a marker of tumour aggressiveness in solid tumours and some haematological malignancies. The prognostic implications of Ki-67 have been examined in numerous well-established studies [4, 5].

A key regulator of the mitochondrial apoptotic pathway is Bcl-2, favouring cell survival by inhibiting adapters necessary for the activation and cleavage of caspases. It promotes cell viability without promoting cell proliferation [6].

Previous studies have described that Bcl-2 is highly expressed in several hematologic and solid malignancies. However, recent evidence suggests that Bcl-2 is an independent favourable prognostic marker in basal cell carcinoma, breast cancer, and non-small cell lung cancer [7–10].

## Aim

This study aimed to investigate the clinicopathological features and immunohistochemical expressions of p53, Ki-67, and Bcl-2 in cutaneous basal cell carcinoma focusing on histological subtypes. Their roles and possible interactions in the development and progression of BCC are discussed.

## Material and methods

### Data and specimen selection

A cross-sectional study was carried out in the Dermatology Service of Civil Hospital of Guadalajara from June 2018 to June 2019, where a total of 50 BCCs from 50 patients from Western Mexico were analysed and grouped by histological subtype. Samples were excluded if there was insufficient material, over-fixed material, or artefact by the process. Clinical data including gender and age at diagnosis were recorded. This study was approved by the local ethics committee and the institutional review board.

### Histopathologic examination

The histopathological subtypes were re-confirmed and in tumours with mixed histological subtypes, the predominant component was recorded. Additional variables such as desmoplasia, Clark level, solar elastosis and pigment deposition were also evaluated. The haematoxylin-eosin stained sections of all samples were reviewed by two experienced pathologists (D.E.V) to determine the histological subtype and then immunohistochemistry expression was graded.

### Immunohistochemistry

Immunohistochemical staining of p53, Ki-67 and Bcl-2 was performed using automated staining with the

Benchmark ULTRA system (Ventana Medical Systems, AZ, USA). For each BCC, a 4 µm thick section of paraffin was mounted on a Menzel SuperFrost Plus adhesive slide and the manufacturer's specifications were followed using the Ventana Ultraview System. The antibodies used were the CONFIRM mouse monoclonal anti-Bcl-2 antibody 124 (Ventana), primary anti-p53 antibody Bp53-11 (Ventana) and CONFIRM rabbit anti-Ki-67 monoclonal primary antibody 30-9 (Ventana).

All slides included positive controls and the omission of the primary antibody was used as a negative control.

### Expression grading

Cytoplasmic positivity for Bcl-2 and nuclear positivity for Ki-67 and p53 were evaluated. Only histological areas that presented the neoplasm were assessed. The staining intensity was classified from 0 to 3 (0 for negative staining, 1 for weak staining, 2 for moderate staining, and 3 for strong staining). Positivity was determined by observing all slides at low power (40×) and then randomly selecting three fields at high power (400×) to estimate an average percentage of immunolabelled positive cells. Disparities in percentages were solved by digital imaging analysis with QuPath software version 0.2.2 [10]. Percentage positivity was ranked from 0 to 4; 0 (no positive cells), 1 (< 10% of positive cells), 2 (10–50% of positive cells), 3 (51–80% of positive cells) and 4 (> 80% of positive cells). The sum of the ordinal scores for percentage and intensity of immunostaining allowed obtaining an expression score from 0 to 7, where 0–3 was considered as a low expression and 4–7 as a high expression.

### Ethics

Institutional ethics committee registration number: 024/20. This study was performed in line with the principles of the Declaration of Helsinki.

### Statistical analysis

The relationship between biomarker expression and the clinicopathological features were analysed using the Fisher's exact test. Multinomial linear regression was conducted to recognize the association between the biomarkers ki-67, p53 and Bcl-2 and the rest of the variables. A correlation test was used to estimate the degree of association between variables. Statistical significance was considered with a value of  $p < 0.05$ . Data were analysed using IBM SPSS® statistics version 25 (Armonk, New York, USA).

### Quality assessment

The Reporting Recommendations for Tumor Marker Prognostic Studies (REMARK) guidelines were used to evaluate manuscripts' quality in order to enhance the possibilities of comparing results across studies involving molecular biomarkers [11].

## Results

### Clinico-pathological data

A total of 50 BCC samples from 50 patients were included, from June 2018 to June 2019. Clinicopathological results are shown in Table 1.

### Biomarker expression

High expression of p53 was found in 30 (60%) samples and low expression in 20 (40%) samples. High expression of ki-67 was found in 33 (66%) samples and low expression in 17 (34%) samples. High expression of Bcl-2 was found in 24 (48%) samples and low expression in 26 (52%) samples.

The immunohistochemical patterns evaluated as low (0–3) and high (4–7) expressions between non-aggressive BCCs and aggressive BCCs are shown in Figure 1. Representative immunohistochemical expressions are shown in Figure 2.

### Relationship and association of biomarker expression and clinicopathological variables

We observed a statistically significant association when BCCs were grouped as non-aggressive and aggressive subtypes for p53 ( $p = 0.04$ ) and Bcl-2 ( $p < 0.01$ ). No statistical association was found between Ki-67 and tumour aggressiveness. No statistical association was found between histological variables and biomarkers (Table 2).

Multinomial logistic regression analysis showed that the expression of Ki-67, p53 and Bcl-2 was statistically associated with the aggressiveness of the tumour. Compared with BCCs with low Bcl-2 expression, BCCs with high expression of Bcl-2 were obviously associated with a non-aggressive subtype (OR = 8.79; 95% CI: 3.21–61.9,  $p = 0.01$ ). In addition, the odds ratio for associating a low expression of p53 (as compared to a high expression of p53) with a non-aggressive BCC was significant (OR = 6.4; 95% CI: 1.17–23.9,  $p = 0.006$ ).

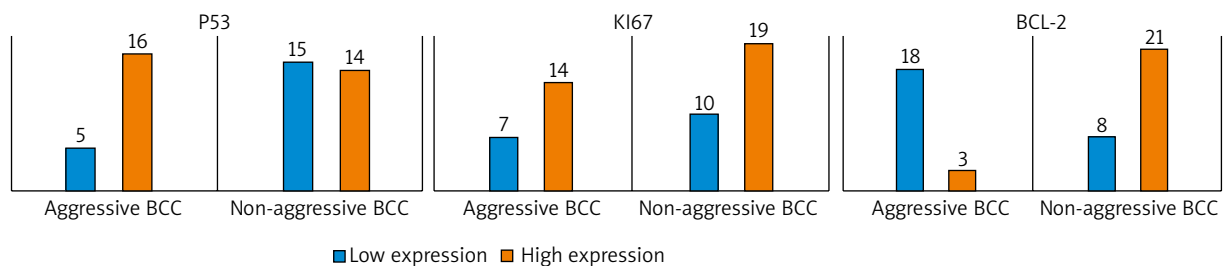
On the other hand, the probability of associating a high level of ki-67 (compared to a low level of ki-67) with a low aggressiveness was significant (OR = 2.96; 95% CI: 0.39–8.78,  $p = 0.05$ ). Given that the latter value presents a borderline  $p$ -value, it should be taken with caution. There was no statistical significance with age, histological subtype, pigment deposition, desmoplasia or Clark level (Table 3).

### Correlation between Bcl-2, p53, and Ki-67 expression and clinicopathological characteristics

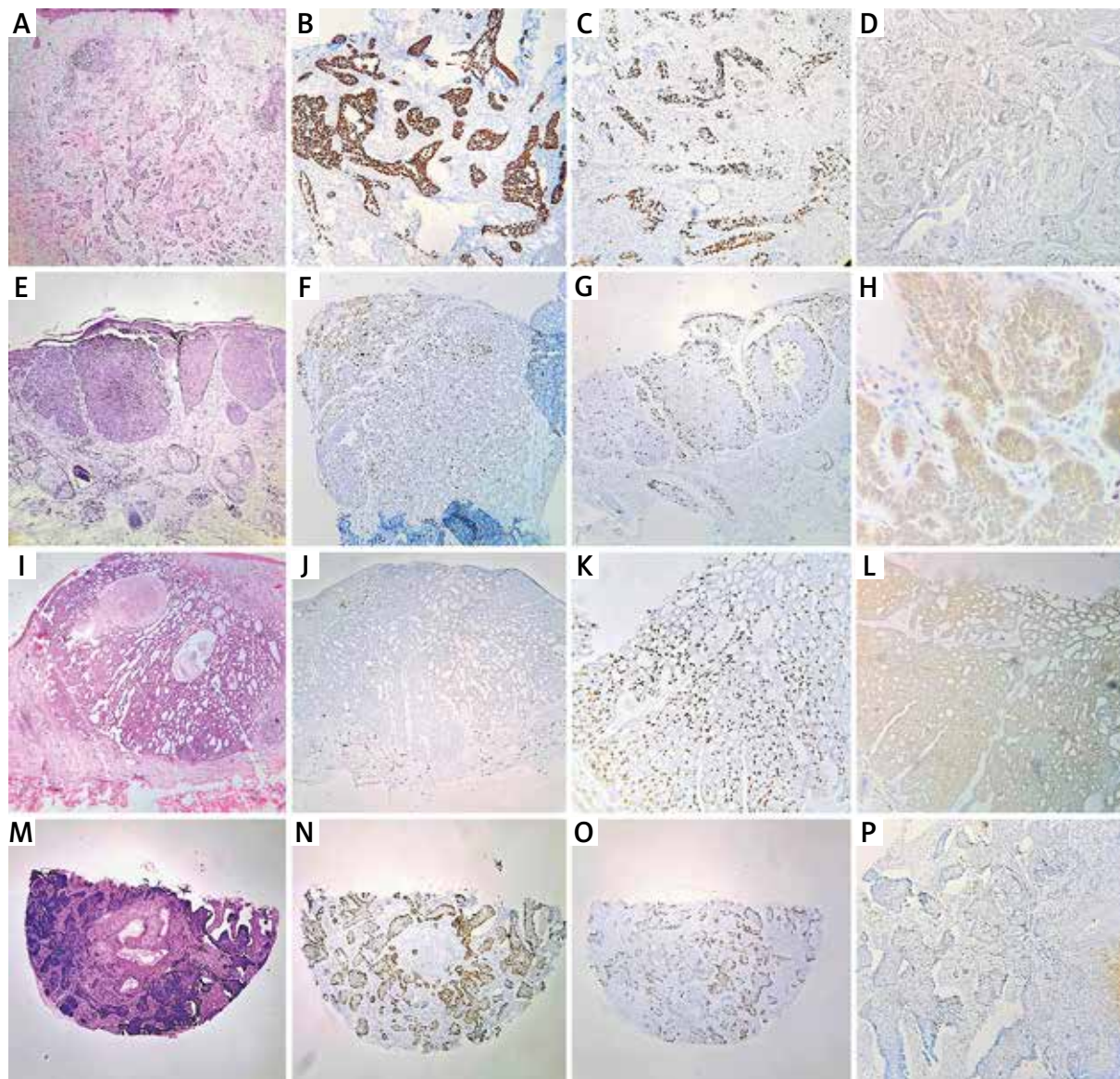
A correlation test among biomarker expression and clinicopathological features was performed. Bcl-2 showed a significant negative correlation with age (correlation coefficient =  $-0.308$ ,  $p \leq 0.05$ ) (Table 4).

**Table 1.** Clinical and histological characteristics ( $n = 50$ )

Variable	BCC n (%)
Age [years]	67 ±15
Gender:	
Male	20 (40)
Female	30 (60)
Histological subtype:	
Nodular	25 (50)
Morpheaform	10 (18)
Micronodular	10 (18)
Adenoid	4 (8)
Metatypical	2 (4)
Solar elastosis:	
Present	39 (78)
Absent	11 (22)
Pigment deposition:	
Present	24 (48)
Absent	26 (52)
Clark level:	
V	5 (10)
IV	27 (54)
III	17 (34)
II	1 (2)
I	0 (0)
Desmoplasia:	
Severe	7 (14)
Moderate	26 (52)
Mild	15 (30)
Absent	2 (4)



**Figure 1.** Immunohistochemical expression patterns in aggressive and non-aggressive (less aggressive) groups



**Figure 2.** Representative images of immunohistochemical expression of p53 (B, F, J, N), Ki-67 (C, G, K, O) and Bcl-2 (D, H, L, P) in infiltrative morpheaform basal cell carcinoma (A), nodular basal cell carcinoma (E), adenoid basal cell carcinoma (I) and infiltrative morpheaform basal cell carcinoma (M). A, E, G, I, J, M, N, O – 40×, B, C, D, F, K, L – 100×, H – 400×

**Table 2.** Relationship of immunostaining results for aggressive and non-aggressive groups of BCCs

Biomarker	Expression	Total	Non-aggressive BCCs	Aggressive BCCs	P-value*
p53	Low	20	15 (75)	5 (25)	0.04
	High	30	14 (46.6)	16 (53.4)	
Ki-67	Low	17	10 (58.8)	7 (41.2)	0.93
	High	33	19 (57.5)	14 (42.5)	
Bcl-2	Low	26	8 (30.7)	18 (69.3)	< 0.01
	High	24	21 (87.5)	3 (12.5)	

\*P-value from the Fisher's exact test.

**Table 3.** Multinomial logistic regression analysis

Low BCL-2a		B	Std. error	Wald	P-value	OR (odds ratio)	95% confidence interval for OR	
							Lower bound	Upper bound
Low Ki-67	Age	-0.04	0.027	2.20	0.13	0.96	0.91	1.01
	Histological subtype	-0.72	0.69	1.09	0.29	0.48	0.13	1.87
	Sola elastosis	1.44	1.14	1.6	0.21	4.2	0.45	36.02
	Pigment deposition	-0.57	0.96	0.35	0.56	0.57	0.086	3.73
	Clark level	-0.46	0.68	0.47	0.49	0.63	0.16	2.39
	Desmoplasia	0.61	0.57	0.65	0.42	1.84	0.42	8.16
	Aggressive BCCs	1.08	0.66	2.68	0.101	2.93	0.81	10.6
High Ki-67	Age	-0.007	0.022	0.11	0.74	0.99	0.95	1.04
	Histological subtype	0.29	0.51	0.33	0.57	1.34	0.49	3.65
	Sola elastosis	0.49	0.94	0.27	0.60	1.63	0.26	10.39
	Pigment deposition	-0.25	0.87	0.092	0.762	0.77	0.14	4.23
	Clark level	0.12	0.57	0.05	0.83	1.13	0.37	3.49
	Desmoplasia	-0.25	0.584	0.18	0.67	0.78	0.25	2.46
	Aggressive BCCs	1.08	0.55	3.8	0.05	2.96	0.39	8.78
Low p53	Age	-0.026	0.025	1.11	0.29	0.97	0.93	1.02
	Histological subtype	0.145	0.591	0.60	0.81	1.16	0.36	3.68
	Sola elastosis	1.08	1.04	1.09	0.29	2.94	0.39	22.36
	Pigment deposition	-0.205	0.95	0.047	0.83	0.82	0.13	5.23
	Clark level	-0.43	0.66	0.43	0.51	0.65	0.18	2.35
	Desmoplasia	-0.019	0.65	0.001	0.97	0.98	0.274	3.52
	Aggressive BCCs	1.86	0.67	7.6	0.006	6.4	1.17	23.9
High p53	Age	-0.013	0.022	0.357	0.550	0.98	0.94	1.03
	Histological subtype	-0.101	0.52	0.037	0.85	0.90	0.32	2.52
	Sola elastosis	0.44	0.99	0.19	0.66	1.55	0.22	10.89
	Pigment deposition	-0.67	0.88	0.58	0.45	0.51	0.09	2.86
	Clark level	0.13	0.58	0.05	0.82	1.14	0.37	3.56
	Desmoplasia	0.105	0.616	0.029	0.87	1.11	0.33	3.72
	Aggressive BCCs	0.63	0.56	1.26	0.26	1.88	0.62	5.69
High bcl2	Age	-0.03	0.025	1.77	0.18	0.97	0.92	1.01
	Histological subtype	-0.07	0.59	0.016	0.89	0.93	0.29	2.93
	Sola elastosis	1.43	1.02	1.98	0.16	4.19	0.57	30.7
	Pigment deposition	-0.68	0.92	0.56	0.46	0.51	0.08	3.04
	Clark level	-0.298	0.64	0.22	0.64	0.74	0.21	2.6
	Desmoplasia	0.149	0.66	0.05	0.82	1.17	0.321	4.48
	Aggressive BCCs	2.64	0.76	12.3	0.01	8.79	3.21	61.9

<sup>a</sup>The reference category. R2 = 0.244 (Cox and Snell).

## Discussion

BCC histological subtypes considered aggressive are particularly more complicated to treat due to subclinical extension, local destruction, and unfavourable biologic

behaviour with higher recurrence rates. In addition, histologic subtypes of BCC that include morpheaform, micronodular, and metatypical patterns, are more likely to metastasize [12, 13].

**Table 4.** Correlation among biomarker expression and clinicopathological features

	Age	Ki-67 biomarker	p53 biomarker	Bcl-2 biomarker	Clark level
Age	–	0.144	0.069	–0.308*	–0.088
Ki-67 biomarker	0.144	–	0.103	–0.071	–0.032
p53 biomarker	–0.069	0.103	–	–0.114	0.137
BCL-2 biomarker	–0.308*	–0.071	–0.114	–	–0.231
Clark level	–0.088	0.157	0.144	–0.236	–

\**P*-value < 0.05.

Increased nuclear staining for mutant p53 reflects a loss of function of p53. In sporadic BCCs, inactivating mutations in the *TP53* gene have been found in 50% of BCCs [14]. Our analysis of p53 expression revealed a statistical association with tumour aggressiveness. These findings are consistent with Oana *et al.* [15] who found that infiltrative BCCs had higher p53 expression in comparison to the nodular subtype ( $p = 0.054$ ). Likewise, Shamsimeyandi *et al.* [3] assessed p53 expression between aBCC and nBCC and found a significantly higher expression in the aggressive groups. In addition, Brito *et al.* [16] reported a higher expression in recurrent BCC and infiltrative BCC than the normal epidermis. Mutations of p53 may also have an impact on BCC treatment. A recent study of p53 expression found that cell lines that displayed mutations of p53 were more resistant to imiquimod-induced apoptosis [17]. Moreover, p53 expression was also associated with BCC resistance to photodynamic therapy (PDT) [18].

In our study, the number of tumour cells in BCC expressing Ki-67 antigen exhibited wide variation, and a high expression was found independent of the histological subtype. These findings are consistent with Shamsimeyandi *et al.* [3] who evaluated an equivalent number of cases amongst most histological subtypes ( $n = 22$  aBCCs vs. 20 nBCCs) and found no difference in Ki-67 expression. However, Khalesi *et al.* [19] evaluated nBCC and found a significantly higher expression of Ki-67 in the superficial subtype compared to the nodular subtype. Interestingly, Yerebakan *et al.* [20] found strong differences ( $p < 0.0001$ ) of expression in recurrent tumours but not between histological subtypes.

Our results concerning Bcl-2 are consistent with previous data, where nBCC had a higher expression. Like our study, all following studies found a significant difference between the two groups, with the expression being the highest in nBCCs and lowest in aBCCs. Ramdial *et al.* [21] reported a low Bcl-2 expression in all of their aBCCs compared to nBCCs ( $p < 0.02$ ). Zagrodnik *et al.* [22] examined recurrent tumours in patients treated with radiotherapy and found a significant correlation between low Bcl-2 expression and aBCCs ( $p = 0.0169$ ) but not with recurrences. Sivrikoz *et al.* [9] included more samples of aBCCs ( $n = 77$ ) than nBCC ( $n = 23$ ). The contrast of Bcl-2 expression in aBCC and nBCC in our study suggests that

they form a complex group of tumours that differ considerably in morphologic and biological behaviour, despite the common origin of these tumours from basal stem cells [23]. In the context of these previous studies, our findings suggest that a high expression of Bcl-2 might be a favourable prognosis factor. This can be explained by the finding that although Bcl-2 inhibits apoptosis it may also slow cell growth [24]. It is known that ultraviolet (UV) radiation induces downregulation of Bcl-2 *in vivo* and *in vitro* [25]. A recent study concluded that apoptosis in BCC does not involve BAX and that the apoptotic activity of BCCs is regulated by either less common members of the BCL2 gene family or a BCL2 gene family independent pathway [26]. The increase in genetic mutations induced by UV or other carcinogens together with a spontaneous or UV-induced downregulation of Bcl-2 may result in aggressive biological behaviour in BCCs. UV chronic exposure could explain our findings of the correlation between older age and lower Bcl-2 expression.

## Conclusions

We have found that a high expression of Bcl-2 and a low p53 expression is associated with more indolent histopathological features with better outcomes. Our results suggest that analysis of p53 and Bcl-2 expression in BCC patients may provide useful prognostic information. However, the clinical implications of these interactions in BCC need to be critically evaluated.

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## Conflict of Interest

The authors declare no conflict of interest.

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